

Schiff Bases Ligands Derived from *o*-Phthalaldehyde and Their Metal Complexes with Cu²⁺ and Ni²⁺: Synthesis, Anti-Breast Cancer and Molecular Docking Study

Adnan Taleb Nasser^{1,2} and Rafid Humaidan Al-Asadi^{1,*}

¹Department of Chemistry, College of Education for Pure Sciences, University of Basra, Basra 61004, Iraq

²Hamdan Girls High School, Education Directorate of Basrah, Ministry of Education, Basrah 61004, Iraq

(*Corresponding author's e-mail: dr.rafid74@yahoo.com)

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Abstract

The Schiff bases and their complexes have an observed biological efficacy, so in the current study it has been prepared, characterized and evaluated the biological activity of some Schiff bases and their metal complexes with copper and nickel and based on ortho-phthalaldehyde as a primary compound. A new Schiff bases ligands and metal complexes $\{[\text{Cu}(\text{L})_n(\text{H}_2\text{O})_2]\text{Cl}_2 \cdot m\text{H}_2\text{O}$ and $[\text{Ni}(\text{L})_n] \cdot m\text{H}_2\text{O}$, where $\text{L} = \text{N}', \text{N}'' - ((1\text{E}, 1'\text{E}) - 1, 2\text{-phenylenebis(methanylylidene)})$ di(benzohydrazide) L^1 , $\text{N}', \text{N}'' - ((1\text{E}, 1'\text{E}) - 1, 2\text{-phenylenebis(methanylylidene)})$ di(isonicotinohydrazide) L^2 , 1,2-bis((E)-(2-(2,4,6-trichlorophenyl)hydrazono)methyl) benzene L^3 ; $n = 1, 2$; $m = 0, 1, 3/2, 5/2$ have been synthesized and elucidated by mass, FT-IR, ¹³C and ¹HNMR, molar conductivity, flame-atomic absorption, magnetic susceptibility, Powder-XRD and TG analysis. The results showed that the L^1 and L^2 ligands behave as a tetra-dentate donor and were associated with metal ions in a molar ratio of 1:1, while L^3 ligand was bi-dentate donor and associated with metal ions in 1:2 molar ratio. In addition, the geometric shapes of the prepared complexes were tetrahedral and square planar for Ni²⁺ and Zn²⁺ complexes, and octahedral for Cu²⁺ complexes. The effect of cellular toxicity in the laboratory has been examined by MTT assay for all compounds against the MCF-7 cancer breast cell line and found to have low efficacy except $[\text{Cu}(\text{L}^3)_2(\text{H}_2\text{O})_2] \text{Cl}_2 \cdot \text{H}_2\text{O}$ (5). The copper complex's molecular docking has been performed with breast cancer proteins using the MOE program, and found to target ER α , CDK6 and EGFR proteins by binding to hydrogen bonds and *pi*-interactions.

Keywords: Schiff base, Metal complexes, Molecular docking, Breast cancer, *o*-Phthalaldehyde, MTT, Breast cancer, MOE program

Introduction

Over a long time, Schiff bases take off consideration since of their application in science, such as; antibacterial, anticorrosion, antifungal, anticancer, antioxidant, and their extraordinary affectability, selectivity, and steadiness toward some metal ions [1-5].

Schiff's bases have an imperative part to play within the improvement of coordination and therapeutic chemistry since they effectively frame for metal complexes molecules [6]. These compounds are played within the field of bioinorganic chemistry and different angles of organo-metallic compounds [7,8]. In addition, they were utilized as electronic sources and photonic and laser components because they possessed non-linear visual properties [9,10].

Schiff bases and their complexes containing the azomethine group (-HC=N-). It is shaped by means of the condensation of the primary amines (RNH₂) and the carbonyl compound. The group (-HC=N-) is mainly appropriate for binding to metal ions through the lone pair of nitrogen atom [11].

Transitional metals complexes containing the Schiff base ligands that are linked through nitrogen and oxygen atoms have been of investigate significance over the past a long time and act as active locales and in this manner catalyze chemical reactions [12,13].

Ortho-phthalaldehyde has been applied as an extensively necessary source material in the synthesis and scientific research of Schiff bases, and it contains 2 carbonyl groups that have the ability to react with

nucleophilic reagents to produce di-azomethine compounds. Many research used *o*-phthalaldehyde as a source of the production of Schiff bases, which were later used as ligands to prepare many coordination metal-complexes [14-16].

Cancer is a category of diseases in which a group of cells leads to uncontrolled growth. The drugs that exist to treat cancer are not good enough, so developing more effective chemotherapy drugs is the main challenge for researchers over the years. Breast cancer is the most common malignant tumor in women worldwide and can be treated in ~70 - 80 % of patients with non-metastatic disease in its early stages. Several studies have shown the susceptibility of Schiff base compounds and their metallic complexes to inhibit breast cancer cells [17,18].

The current study focuses on synthesis and identification of some Schiff bases compounds which contains di-azomethine group and their coordination complexes with Copper (II) and Nickel (II) and the study of their effect as anti-breast cancer as well as the study of molecular docking of molecules that have shown effectiveness to find out where to interact with cancer cell proteins.

Materials and methods

The chemicals used in the current study were obtained from well-known companies and used without purification after confirming their melting points. Infrared spectra recorded using Shimadzu model IR Affinity-1 device, using KBr discs between 4,000 - 400 cm^{-1} . The ^1H and ^{13}C NMR spectra were recorded using Bruker 500 MHz spectrometer with DMSO-*d*₆ acting as a solvent. EI technique used to record compounds mass spectra with Agilent Technologies 70 eV spectrometer. TG and DTG measurements under nitrogen were made within a range 25 - 750 °C using the SDT Q600 V20.9 Build 20. The magnetic susceptibility of metal complexes was measured by a Sherwood-scientific magnetic balance at room temperature and using powder samples. Molar conductance of metal-complexes was measured at room temperature in DMSO solvent using HI 2315 Conductivity Meter, and with a concentration (1×10^{-3} M). The percentage of Cu(II) and Ni(II) ions was estimated using the flame atomic absorption technique, and using Sense AA GBC Scientific Equipment device. Powder XRD patterns recorded by Philips X pertpro MPD diffractometer with using Cu K as source, $\lambda = 1.5406 \text{ \AA}$ and Ni as a filter.

Synthesis of ligands

$\text{N}', \text{N}''\text{-(1E,1'E)-1,2-phenylenebis(methanylylidene) di(benzohydrazide) L}^1$

Solution (10 mmol) 1.36 g of benzohydrazide in 10 mL EtOH absolute gradually added to a solution (10 mmol) 1.34 g of *o*-phthalaldehyde in 10 mL EtOH absolute with a few drops of glacial HAc. The mixture was refluxed with stirrer for 2 h. and followed by TLC (ethyl acetate: chloroform, 4:1). The result was cold, filtered and re-crystallized with absolute ethanol. The product was yellow solid crystal. Yield: 84 %, mp: 170 - 172 °C. IR spectrum, ν , cm^{-1} : 3,414 and 3,261 (N-H), 3,064 (Ar-CH), 1,656 (C=O), 1,606 (C=C), 15,62(C=N). ^1H NMR spectrum, δ , ppm: 7.96 - 7.39 m (7H, Ar-H), 8.96 s (1H, CH=N), 12.05 s (1H, NH). ^{13}C NMR spectrum, δ_{C} , ppm: 120.34 (C_{8+10}), 123.04(C_1), 128.20(C_2), 130.37(C_9), 132.43(C_1), 133.30(C_2+C_9), 133.78(C_3+C_6), 146.75(C_4), 163.70(C_5). MS (EI, m/z (%)): 370[M^+ , 0.8].

$\text{N}', \text{N}''\text{-(1E,1'E)-1,2-phenylenebis(methanylylidene) di(isonicotinohydrazide) L}^2$

A similar procedure is used as in L^1 , but used (10 mmol) 1.37 g of isonicotinohydrazide instead of benzohydrazide. The product was a yellow-white crystal. Yield: 89 %, mp: 167 - 168 °C. IR spectrum, ν , cm^{-1} : 3,414 and 3,238 (N-H), 3,070 and 3,049 (Ar-CH), 1,662 (C=O), 1,616(C=C), 1,552(C=N). ^1H NMR spectrum, δ , ppm: 6.68 d (2H, Ar- H_b+H_b), 6.79 d (1H, Ar- H_d), 7.49 - 7.20 m (3H, Ar-H), 7.93 s (1H, CH=N), 9.30 s (1H, NH). ^{13}C NMR spectrum, δ_{C} , ppm: 113.08(C_{7+10}), 119.86(C_1+C_2), 122.80(C_3), 129.25(C_8+C_9), 134.63(C_6), 148.53(C_4), 165.52(C_5). MS (EI, m/z (%)): 371[M^++1 , 10].

1,2-bis((E)-(2-(2,4,6-trichlorophenyl) hydrazono) methyl) benzene L^3

A similar procedure is used as in L^1 , but used (10 mmol) 2.11 g of (2,4,6-trichlorophenyl) hydrazine instead of benzohydrazide. The product was a yellow crystal. Yield: 82 %, mp: 179 - 180 °C. IR spectrum, ν , cm^{-1} : 3,321 (N-H), 3,078 (Ar-CH), 2,860 (Alph-CH), 1,590(C=C), 1,554(C=N). ^1H NMR spectrum, δ , ppm: 7.34 d (1H, H_c), 7.64 s (2H, Ar- H_b+H_b), 7.76 d (Ar-1H, H_a), 8.46 s (1H, CH=N), 9.67 s (1H, NH). ^{13}C NMR spectrum, δ_{C} , ppm: 127.06(C_1), 127.57(C_{7+9}), 127.82(C_2), 128.85(C_3) 129.38(C_6+C_{10}), 132.85(C_8), 137.71(C_5), 138.63(C_4). MS (EI, m/z (%)): 521[M^+ , 0.9].

Synthesis of metal complexes

General procedure

The hot methanolic solution of (1 mmol) of L^1 or L^2 or (2 mmol) of L^3 ligands were gradually added to (1 mmol) of hot methanolic solution of metal chloride (Metal = Cu(II) or Ni(II)) with stirring and

refluxing for 6 - 8 h. The precipitated products were filtered off, washed with water, methanol and diethyl ether. Then it dried and was obtained;

[Cu(L¹)(H₂O)₂]Cl₂ (1): Dark yellow colour. Yield: 76 %, mp: 249 °C dec. IR spectrum, ν , cm⁻¹: 3,477 (O–H), 3,414 (N–H), 3,070 and 3,045 (Ar–CH), 1,616(C=C), 1,573(C=N). Found, %: Cu 11.73. C₂₂H₂₄Cl₂CuN₄O₄. Calculated, %: Cu 11.69. μ_{eff} (B.M): 1.88. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 73.3.

[Ni(L¹)]₂.H₂O (2): Red colour. Yield: 82 %, mp: > 300 °C. IR spectrum, ν , cm⁻¹: 3,444 (O–H), 3,387 (N–H), 3,062 (Ar–CH), 1,591(C=C), 1,554(C=N). Found, %: Ni 14.21. C₂₂H₂₂N₄NiO₃. Calculated, %: Ni 13.14. μ_{eff} (B.M): 2.84. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 3.3.

[Cu(L²)(H₂O)₂] Cl₂ (3): Green colour. Yield: 68 %, mp: > 300 °C. IR spectrum, ν , cm⁻¹: 3,514 (O–H), 3,446 and 3,363 (N–H), 3,086 and 3,057 (Ar–CH), 1,645 (C=C), 1,608 (C=N). Found, %: Cu 11.69. C₂₂H₂₂Cl₂CuN₆O₄. Calculated, %: Cu 11.67. μ_{eff} (B.M): 1.91. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 63.7.

[Ni(L²)]₂.5/2H₂O (4): Yellow colour. Yield: 80 %, mp: > 300 °C. IR spectrum, ν , cm⁻¹: 3,442 (O–H), 3,390 (N–H), 1,608(C=C), 1,566(C=N). Found, %: Ni 14.56. C₂₀H₂₃N₆NiO_{4.5}. Calculated, %: Ni 12.27. μ_{eff} (B.M): 3.10. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 4.2.

[Cu(L³)₂(H₂O)₂] Cl₂.H₂O (5): Yellow colour. Yield: 59 %, mp: > 300 °C. IR spectrum, ν , cm⁻¹: 3,221 (O–H), 3,190 (N–H), 3,078 (Ar–CH), 2,880 (Alph–CH), 1,585(C=C), 1,539(C=N). Found, %: Cu 5.31. C₄₀H₃₀Cl₁₄CuN₈O₃. Calculated, %: Cu 5.16. μ_{eff} (B.M): 1.76. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 76.1.

[Ni(L³)₂].3/2H₂O (6): White colour. Yield: 45 %, mp: 277 °C dec. IR spectrum, ν , cm⁻¹: 3415 (O–H), 3,323 and 3,302 (N–H), 3,078 and 3,047 (Ar–CH), 2,860 (Alph–CH), 1,581(C=C), 1,556(C=N). Found, %: Ni 4.91. C₄₀H₂₇Cl₁₂N₈NiO_{1.5}. Calculated, %: Ni 5.20. μ_{eff} (B.M): 0.89. Molar conductivity (DMSO, Ohm⁻¹cm²mol⁻¹): 15.2.

Anti-cancer activity

MTT assay [19] was employed to test *in-vitro* cytotoxicity of studied compounds. Human breast cancer (MCF-7 cell line) was obtained from National Cell Bank of Iran. Cells grew in RPMI-1640 (Gibco) and DMEM medium (Gibco), respectively, with 10 % FBS (Gibco) supplemented with antibiotic (100 U/mL penicillin and 100 µg/mL streptomycin). Cells were kept at 37 °C under wet air containing 5 % CO₂ and passed utilizing trypsin/EDTA (Gibco) and phosphate- buffered saline solution (PBS). The cultured media and conditions used for cell culture as 3D colonies were the same as single-layer cell culture.

Molecular docking

Molecular docking study of compound 5 was conducted using MOE (Molecular Operating Environment) 2019. The crystal structure of proteins was obtained from Protein Data Bank (PDB) (<https://www.rcsb.org/> structure). Processing of study compound, proteins and docking procedures was carried out in accordance with literature [20].

Results and discussion

The condensation reaction between *o*-phthalaldehyde and some hydrazine derivatives at a 1:2 mole ratio gave Schiff bases ligands L¹, L² and L³, **Figure 1** (i). Reaction of L¹ and L² ligands with Cu(II) and Ni(II) ions in methanol gave the complexes (1 - 4) by a 1:1 ratio, **Figure 1** (ii) a, while reaction of L³ ligands with Cu(II) and Ni(II) ions gave the complexes (5,6) with 1:2 mole ratio, **Figure 1** (ii) b.

Characterization methods

The proposed structure of ligands was created by mass, infrared and nuclear magnetic resonance techniques. Mass spectra of prepared ligands showed molecular ion peak at $m/z = 370$, 371 and 521 for L¹, L² and L³, respectively, but with low relative abundance, and the appearance of a set of fragments which indicating the presence of the proposed compound. The mass spectra of complexes did not exhibit the peaks that due to their molecular ions. However, it has shown the fragments confirming the presence of these compounds.

The FT-IR spectra for synthesis compounds showed weak bands at range 3,446 - 3,238 cm⁻¹ which assign to stretching vibration of –NH groups, and 1 or 2 bands at the range between 3,045 - 3,086 cm⁻¹ due to symmetrical and asymmetrical stretching vibration of C - H aromatic. In addition to, the strong band at range 1,608 - 1,552 cm⁻¹ attributed to stretching vibration of azomethine group, this band has a shifting in complexes compounds due to the coordination between metals and ligands. The L¹ and L² compounds were characterized by the appearance of a strong band at 1,656 and 1,662 cm⁻¹ due to the

presence of the carbonyl group, this band has disappeared in the coordination complexes, it is evidence the participation of C=O group in the coordinate process. The spectra of complexes were also characterized by the appearance of the stretching vibration of OH group as a broad band in the range $3,221 - 3,514 \text{ cm}^{-1}$ because it contained the coordinate and lattice water molecules.

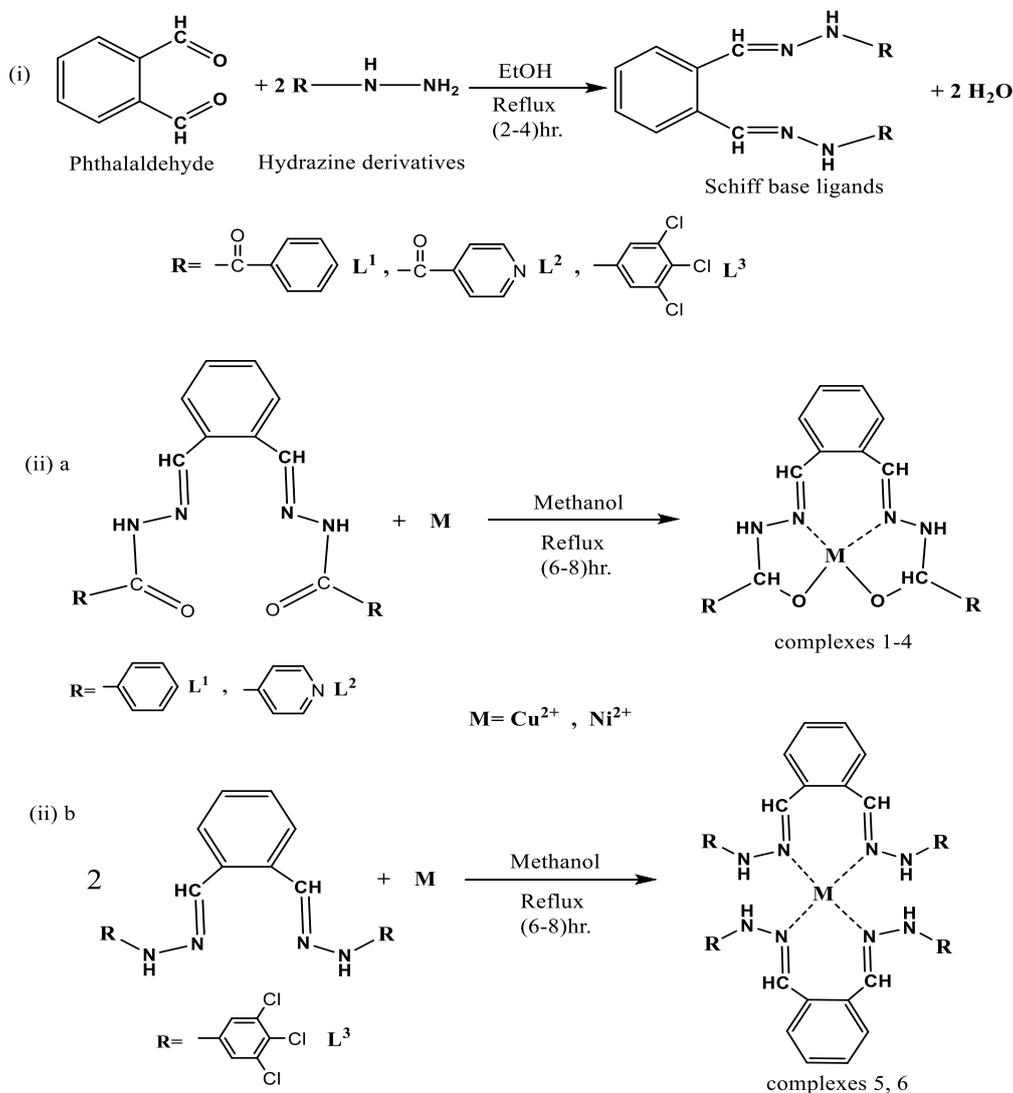


Figure1 Preparation pathway to ligands and metal complexes.

^1H NMR spectra of ligands, **Figure 2**, demonstrated a singlet, doublet and multiplet signals at the range 6.68 - 7.96 ppm that can be attributed to aromatic protons. The spectra confirmed the presence of the azomethine group with the appearance of a singlet signal at 8.96, 7.93 and 8.46 ppm for L^1 , L^2 and L^3 , respectively, while proton of amide group (-NH) appeared as a singlet signal at the 12.05, 9.30 and 9.67 ppm. ^{13}C NMR spectra have proven the correct skeletal structure of the prepared ligands. The signals at 163.70 ppm for L^1 and 165.52 ppm for L^2 assign to carbons of C=O group, while the signals at 146.75, 148.53 and 138.63 ppm for L^1 , L^2 and L^3 were due to carbon of azomethine group. Aromatic carbons appeared in the range of 113.08 - 137.71ppm.

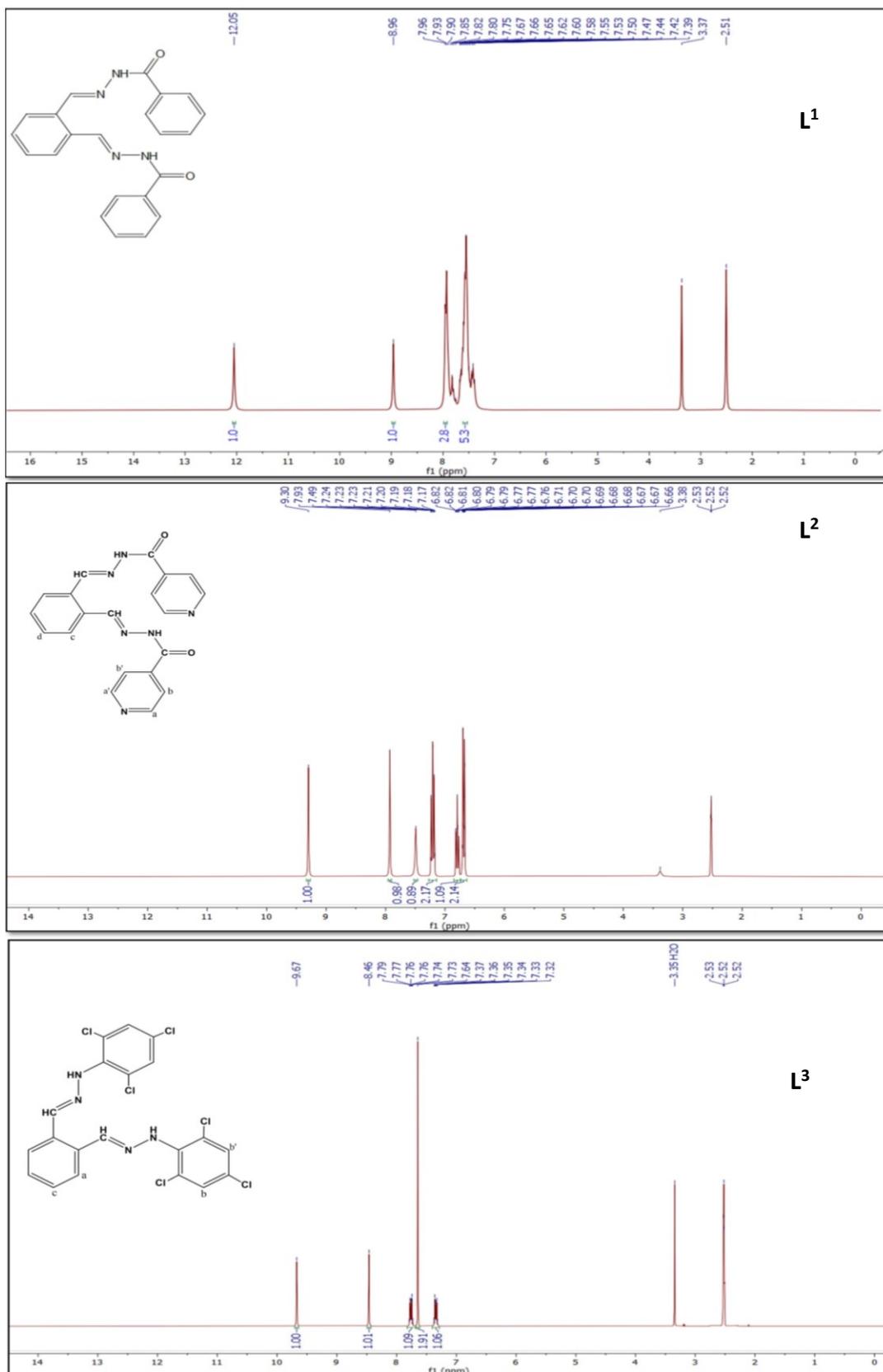


Figure 2 ¹H NMR spectra of ligands L¹, L² and L³.

The metal: Ligand mole ratio of complexes determined by flame-atomic absorption, it was found that the Cu(II) and Ni(II) ions formed 1:1 chelates with L¹ and L² ligands, while was formed 1:2 chelates with L³. The molar conductivity values of Cu-complexes were found to be located in the range of 63.7 - 76.1 Ohm⁻¹ cm² mol⁻¹, where they are predicted to possess electrolyte properties and containing a counter-ion, while Ni-complexes were conductive values within range 3.3 - 15.2 Ohm⁻¹ cm² mol⁻¹, which confirm that they did not contain counter ions with non-electrolyte properties [21].

The values of effective magnetic moment (μ_{eff}) of Cu-complexes, 1, 3 and 5, were at range 1.76 - 1.91 B.M, which indicates that they contain 1 unpaired electron in *d*-orbital and not found the participation of the orbitals magnetic moment, so the suggested geometric shape around copper ion in these complexes is octahedral (sp³d²). Ni-Complexes, 2 and 4, μ_{eff} value was (3.3 and 4.2 B.M), respectively, that this data indicates the presence of 2 unpaired electrons in *d*-orbital (high spin system), in addition to the presence of the orbitals moment contribution, so the expected geometric shape of these complexes is tetrahedral (sp³), but complex 6, the value of μ_{eff} was 0.89 B.M, these value refers that it has diamagnetic (low spin system) and that its geometric shape is a square planar (dsp²) [22].

Figure 3 and **Table 1** show the stages of thermal decomposition of metal complexes, the decomposition stages and thermograms refer the presence of lattice and coordinated water in structure of complexes molecules, which decompose in the early stages of disintegration at range 50 - 272 °C, while the other stages included the loss of parts of the ligands.

Table 1 Thermogravimetric analysis data of metal complexes.

Complexes	Stages	Temp. range (°C)	Decomposition parts	Weight loss %	
				Calculated	Found
[Cu(L ¹)(H ₂ O) ₂]Cl ₂	1	50 - 272	2H ₂ O	6.65	6.37
	2	272 - 345	Cl ₂	13.12	12.95
	3	345 - 394	CO ₂	8.13	7.67
[Ni(L ¹)]·H ₂ O	1	50 - 145	H ₂ O	4.02	3.09
	2	145 - 293	NH	3.35	3.31
	3	293 - 362	C ₇ H ₇ N	23.48	23.93
	4	362 - 475	C ₇ H ₇	20.35	20.03
[Cu(L ²)(H ₂ O) ₂]Cl ₂	1	50 - 242	2H ₂ O+CO ₂	14.73	11.58
	2	242 - 292	CH ₂ N ₂	8.28	7.91
	3	292 - 378	C ₈ H ₆	20.11	20.47
[Ni(L ²)]·5/2H ₂ O	1	50 - 214	5/2H ₂ O	9.45	9.21
	2	214 - 395	2NH+CO ₂	15.96	15.87
	3	395 - 475	C ₈ H ₄	21.00	20.80
[Cu(L ³) ₂ (H ₂ O)]Cl ₂ ·H ₂ O	1	50 - 108	3H ₂ O	4.38	4.08
	2	108 - 172	C ₈ H ₆	8.28	8.22
	3	172 - 228	C ₈ H ₇ N	9.51	9.97
[Ni(L ³) ₂]·3/2H ₂ O	1	50 - 206	3/2H ₂ O	2.39	2.40
	2	205 - 672	C ₂₅ H ₁₂ N ₂	42.47	42.30

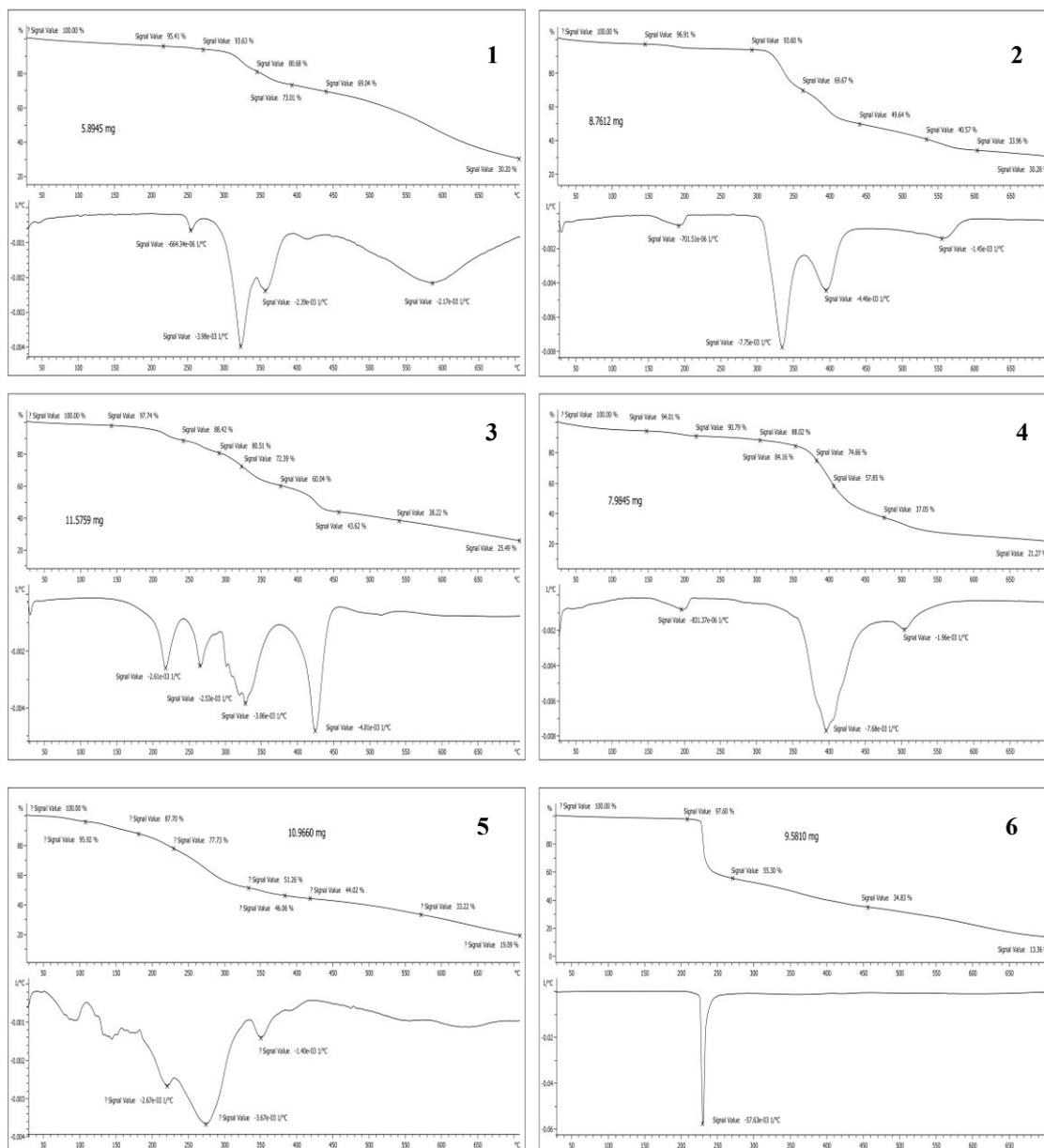


Figure 3 TG Patterns of complexes $[\text{Cu}(\text{L}^1)(\text{H}_2\text{O})_2]\text{Cl}_2(1)$, $[\text{Ni}(\text{L}^1)]\cdot\text{H}_2\text{O}(2)$, $[\text{Cu}(\text{L}^2)(\text{H}_2\text{O})_2]\text{Cl}_2(3)$, $[\text{Ni}(\text{L}^2)]\cdot 5/2\text{H}_2\text{O}(4)$, $[\text{Cu}(\text{L}^3)_2(\text{H}_2\text{O})_2]\text{Cl}_2\cdot\text{H}_2\text{O}(5)$ and $[\text{Ni}(\text{L}^3)_2]\cdot 3/2\text{H}_2\text{O}(6)$.

The powder XRD patterns of complexes, **Figure 4** recorded at the range $2\theta = 10 - 90^\circ$. Copper complexes, 1 and 3 have crystalline shapes, each showed 4 crystalline peaks at $2\theta = 10.91^\circ$, 11.73° , 15.71° and 16.49° for complex 1, and $2\theta = 13.15^\circ$, 17.64° , 22.75° and 26.01° for complex 3. Nickel complexes, 2 and 4 showed a semi-crystalline character through low-split peaks at $2\theta = 22.17^\circ$, 22.85° and 29.29° for complex 2, and $2\theta = 10.68^\circ$, 19.63° and 21.76° for complex 4. While the L^3 ligand complexes (5 and 6) have shown amorphous properties. The crystallite size of complexes was calculated in nanometer units using the Debye-Scherrer equation [23], and found that it was equal to 37.89, 54.90, 48.54 and 31.56 nm for the complexes 1, 2, 3 and 4, respectively.

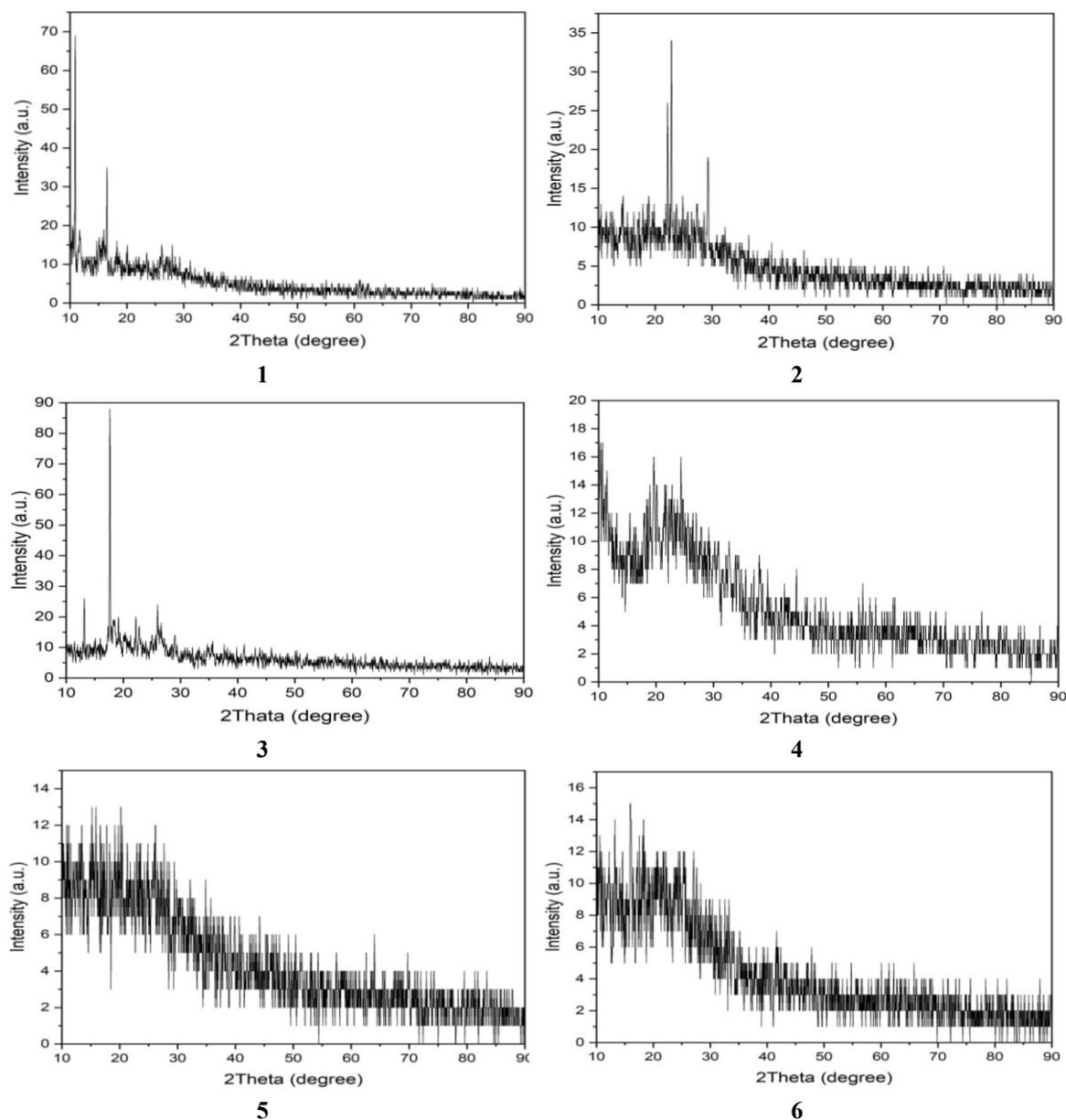


Figure 4 XRD Patterns of complexes $[\text{Cu}(\text{L}^1)(\text{H}_2\text{O})_2]\text{Cl}_2$ (1), $[\text{Ni}(\text{L}^1)].\text{H}_2\text{O}$ (2), $[\text{Cu}(\text{L}^2)(\text{H}_2\text{O})_2]\text{Cl}_2$ (3), $[\text{Ni}(\text{L}^2)].5/2\text{H}_2\text{O}$ (4), $[\text{Cu}(\text{L}^3)_2(\text{H}_2\text{O})_2]\text{Cl}_2.\text{H}_2\text{O}$ (5) and $[\text{Ni}(\text{L}^3)_2].3/2\text{H}_2\text{O}$ (6).

Anticancer activity

The activity of compounds against breast cancer cells line (MCF-7) was studied using 5 different concentrations (31.25, 62.5, 125, 250, 500 $\mu\text{g}/\text{mL}$) per compounds and the effectiveness of inhibition was calculated at each concentration, in addition to calculating the concentration values of the maximum half effect IC_{50} . All synthesis compounds demonstrated weak anti-cancer activity with IC_{50} values above 70 $\mu\text{g}/\text{mL}$, compared with IC_{50} value of cisplatin as a standard compound ($\text{IC}_{50}=1.5 \mu\text{g}/\text{mL}$) [24]. But 1 compound, complex 5 showed a moderated activity, $\text{IC}_{50}=8.0 \mu\text{g}/\text{mL}$ with 52.16 % inhibition ratio at the lowest concentration, **Table 2**, which may be due to the presence of chlorine and copper atoms in the structure of complexes.

Table 2 Inhibition ratio of MCF-7 cell line and IC50 values for studied compounds.

Compounds	Conc.($\mu\text{g/mL}$)					IC ₅₀ ($\mu\text{g/mL}$)
	31.25	62.5	125	250	500	
	Cells inhibition (%)					
L ¹	4.73	24.72	32.29	32.46	33.91	1612.3
L ²	22.14	35.52	55.02	58.09	73.89	132.4
L ³	17.79	31.00	43.26	75.34	75.82	132.5
[Cu(L ¹)(H ₂ O) ₂]Cl ₂	25.69	39.87	51.48	56.64	76.30	125.1
[Ni(L ¹)] ₂ .H ₂ O	17.14	51.80	56.64	63.89	65.18	117.6
[Cu(L ²)(H ₂ O) ₂]Cl ₂	17.31	32.69	42.55	54.09	55.76	76.3
[Ni(L ²)] ₂ .5/2H ₂ O	25.96	39.01	54.48	56.64	75.30	125.4
[Cu(L ³) ₂ (H ₂ O)]Cl ₂ .H ₂ O	52.16	63.22	63.94	71.87	72.32	8.0
[Ni(L ³) ₂].3/2H ₂ O	1.67	9.24	21.01	31.33	64.21	423.7

Molecular docking study

The study of the molecular docking of compound 5 was conducted against breast cancer proteins, MCF-7 cell line which are: ER α (PDP: 3ERT), PR (PDP: 4OAR), EGFR (PDP: 2J6M), mTOR (PDP: 4DRH), CDK2 (PDP: 4FX3), CDK6 (PDP: 3NUP) and Akt (PDP: 5KCV) [25-27]. It was the strongest interaction with 3ERT, 3NUP and 2J6M proteins where it gave the highest affinity energy (S) and the lowest RMSD values [28], **Table 3**. Docking analysis is used to determine how amino acid residues and hydrogen bonds of target proteins interact with compound 5. 2D and 3D forms (**Figure 4**) show a 1 *pi*-interaction of the hydrophobic phenyl moiety with the amino acid Cys-530 of protein 3ERT and 2 hydrogen bonds with 3NUP protein, the first with Lys-129 at a distance of 2.43 Å and other with Asp-102 at a distance of 2.6 Å. While the interaction with protein 2J6M was by forming a hydrogen bond with amino acid Lys-875 with a distance of 2.56 Å, in addition to forming a *pi*-interaction of phenyl ring with amino acid phe-723.

Table 3 Molecular docking data of compound 5 with target 3ERT, 3NUP and 2J6M proteins.

Proteins (Receptors)	RMSD	Affinity energy (S) Kcal/mol	Interaction		
			Type	Amino acid	H-Bonding distance(Å)
3ERT	1.727	-7.432	<i>pi</i> -interaction	Cys530	-
3NUP	1.638	-7.360	H-Bonding	Lys129	2.43
			H-Bonding	Asp102	2.67
2J6M	1.9001	-7.351	H-Bonding	Lys875	2.56
			<i>pi</i> -interaction	Phe723	-

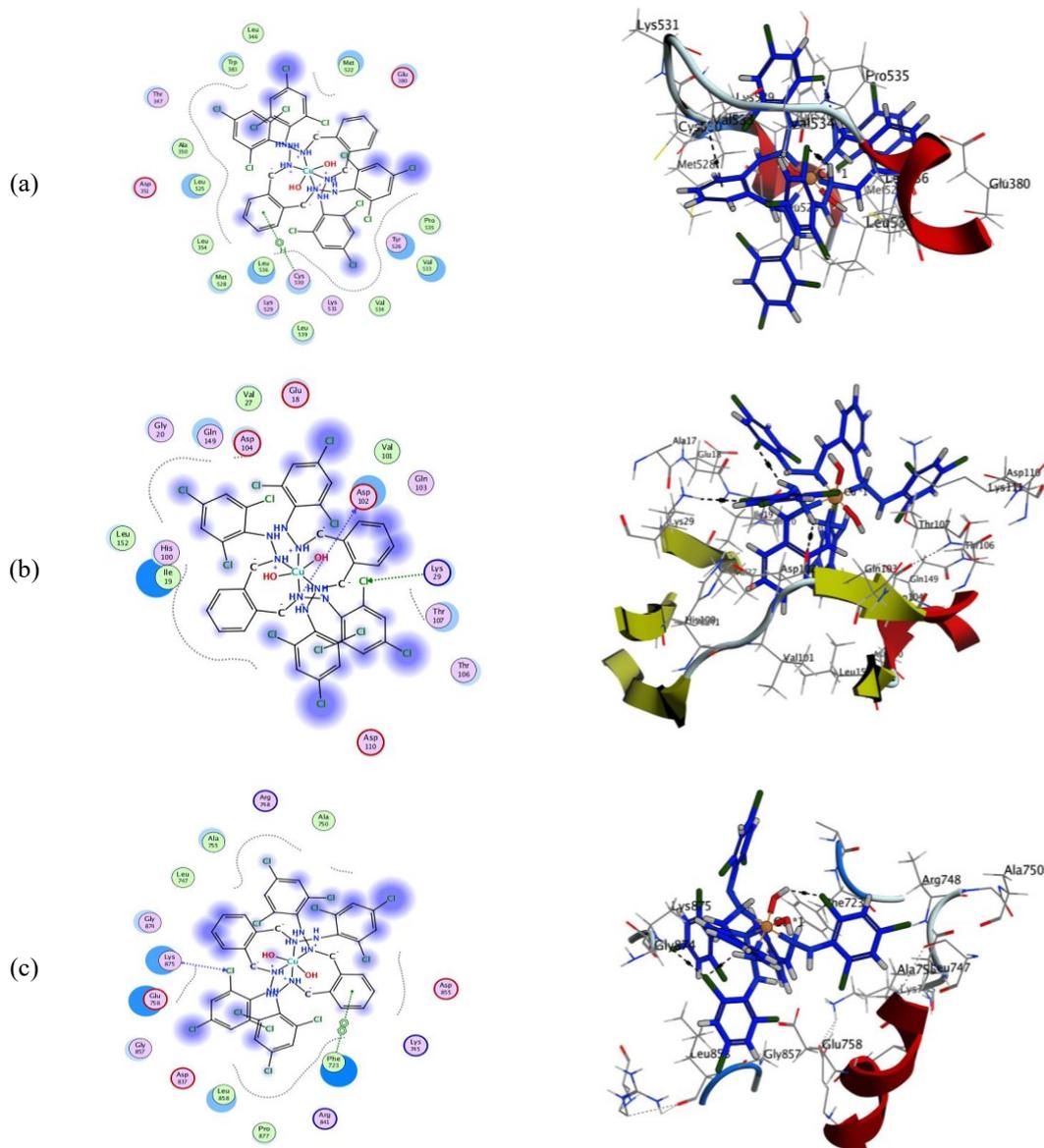


Figure 4 2D and 3D forms of compound 5 with target proteins; (a): 3ERT, (b): 3NUP and (c): 2J6M.

Conclusions

Characterization techniques have validated the proposed structure of the synthesized compounds. L^1 and L^2 acted with Cu^{2+} and Ni^{2+} as a tetra dentate ligands type N_2O_2 , while the L^3 was a bi-dentate ligand type N_2 . In addition, the geometry of complexes was octahedral (sp^3d^2) with Cu^{2+} and tetrahedral (sp^3) and square planar (dsp^2) with Ni^{2+} . L^1 and L^2 complexes with copper had crystalline properties and with nickel were semi-crystalline, while the L^3 complexes have amorphous characters. The biological study proved that copper complex with L^3 is the only one that is effective against breast cancer (MFC-3 cell line) and it targets $ER\alpha$, CDK6 and EGFR proteins.

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