

## Effect of *Sonneratia alba* Fruit Extract on LOX-1 and Nrf2 Expression in the Aortic Arches of High Fat Diet Induced Rats

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### Abstract

Atherosclerosis, the primary cause of ischemic heart disease, is a major contributor to global mortality. This highlights the urgent need for therapeutic strategies that are not only beneficial and safe but also economically sustainable, such as those derived from natural bioactive compounds. This study investigated the anti-atherogenic effect of mangrove *Sonneratia alba*, a source of bioactive compounds with known antioxidant and anti-inflammatory properties. The objective was to determine the extract's effect on the pro-atherogenic receptor LOX-1 and its subsequent effect on foam cell formation and the endogenous antioxidant axis Nrf2-MnSOD in the aortic arch of male Wistar rats exposed to a 10-week high-fat diet. A total of twenty-four rats were randomly divided into six groups; normal control, HFD control, atorvastatin control (10 mg/day) and HFD groups treated with *S. alba* extract at 200, 400 and 800 mg/kg/day. The expression of LOX-1, Nrf2, and MnSOD was assessed using immunohistochemistry, while foam cells were identified through H&E staining. The results showed *S. alba* fruit extract at the dose of 800 mg/kg/day significantly reduced circulating LDL-C levels. Within the aortic arch, the *S. alba* extract at the same dose significantly suppressed LOX-1 expression, significantly reduced approximately 82% of foam cell formation and significantly elevated Nrf2 expression. The expression of MnSOD was enhanced however this change did not reach statistical significance. These data collectively suggest that *S. alba* fruit extract ameliorates early atherogenic features, associated with the modulation of key regulatory proteins LOX-1 and Nrf2, and subsequent reduction in foam cell formation. Thus, the extract seems promising as a natural agent for vascular protection, but further functional investigations are needed to definitively confirm the underlying mechanism.

**Keywords:** Atherosclerosis, Antioxidant, Foam cells, Hyperlipidemia, LOX-1, Nrf2, MnSOD, *Sonneratia alba*

### Introduction

Atherosclerosis, a chronic inflammatory disease of the arterial wall is the primary pathological basis for ischemic heart disease, that persists the leading cause of morbidity and mortality globally [1]. Despite the availability and efficacy of atherosclerosis therapies,

particularly those focused in lipid control, the global incidence of this disease continues to rise [2]. Furthermore, the mortality rate associated with this disease has been reported to increase from 5.367 million in 1990 to 8.992 million in 2021 [3]. The early stages of

atherosclerosis are annotated by endothelial dysfunction, retention of low-density lipoprotein (LDL) in the sub endothelial space and its subsequent modification, particularly through oxidation. Oxidized low-density lipoprotein (oxLDL) serve as a critical driver of the disease's progression [4]. It is recognized by scavenger receptors on macrophages, most notably the lectin-like oxidized low-density lipoprotein receptor-1 (LOX-1), that modulates the uncontrolled uptake of lipids and drives the transformation of macrophages into foam cells. The accumulation of foam cells in the arterial intima is a hallmark lesion of early atherogenesis [5]. Therefore, interrupting the oxLDL/LOX-1 pathway and mitigating foam cells growth represents one of major therapeutic strategies against atherosclerosis [6]. Despite numerous studies targeting LOX-1 have been conducted using either natural or synthetic compounds, research in this area remains limited and requires further investigation regarding its efficacy and safety [7].

Oxidative stress plays a pivotal role in this cascade, not only by generating oxLDL but also by overwhelming the intrinsic antioxidant defenses of the vascular wall [8]. The excessive production of reactive oxygen species (ROS) driven by enzymes like NADPH oxidase (NOX) and mitochondrial dysfunction is what fundamentally causes the oxidation of LDL [9]. Macrophages via its scavenger receptors uptake the oxidized LDL, turning them into lipid laden foam cells that accumulate in arterial walls. This process triggers inflammation and endothelial dysfunction, recruits more immune cells, induces vascular smooth muscle cells phenotype switching and migration which leads to plaque formation and vicious cycle of oxidative stress [10]. A key cellular defense mechanism against oxidative stress is the nuclear factor erythroid 2-related factor 2 (Nrf2) signaling pathway. Under normal condition, Nrf2 is sequestered in the cytoplasm; however, upon exposure to ROS, it is translocated into the nucleus. There, Nrf2 binds to the antioxidant response element (ARE) and induces the expression of numerous cytoprotective genes, including antioxidant enzymes such as manganese superoxide dismutase (MnSOD) [11]. Activating the Nrf2 pathway is hypothesized to attenuate the pro-atherogenic environment by neutralizing ROS and potentially inhibiting the formulation of oxLDL [12]. Thus, Nrf2

represents an effective therapeutic target for atherosclerosis, complementing the existing anti-cholesterol therapies [13].

Natural product, especially those derived from medicinal plants, serve as a rich source of bioactive compounds with potential anti-atherosclerotic effects, often related to their strong antioxidant and anti-inflammatory activities [14]. *Sonneratia alba* (L.f.) Sm., a widely distributed mangrove species, is traditionally used for its therapeutic benefits. Phytochemical analyses reveal that its fruits are particularly rich in polyphenols, flavonoids, triterpenoid and steroids that are well known for their potent radical-scavenging activities [15,16]. Other studies have identified several important phytoconstituents in *S. alba*, including oleanolic acid, ursolic acid,  $\alpha$ - dan  $\beta$ - amyrin cinnamate, stigmasterol,  $\beta$ -stigmasterol, lupeol, and squalene [15,16]. Oleanolic acid is recognized significant capability to lower total cholesterol, triglyceride and LDL-C levels, and increase high-density lipoprotein cholesterol (HDL-C) levels, prevent aortic endothelial thickening and inhibit atherosclerotic plaque formation in high fat diet animal model [17,18]. Moreover, it inhibits type 1 collagen formation and enhance the expression of Nrf2 and heme-oxygenase 1 (HO-1) in LOX-1 knockout human umbilical vein endothelial cells (HUVEC) that is induced by oxLDL [19]. Lupeol, another identified compound, demonstrates antioxidant properties and significantly elevates nitric oxide (NO), superoxide dismutase (SOD), catalase and glutathione (GSH) in cardiac injury animal model [20], promotes macrophages polarization to an inflammatory phenotype, thereby preventing inflammatory signal transmission and diminishing intracellular lipid accumulation, subsequently prevents atheroma plaque formation [21]. Lupeol seems to protect against myocardial ischemia-reperfusion injury by regulating nuclear factor kappa B (NF- $\kappa$ B) and Nrf2 signaling pathways [22]. Stigmasterol significantly suppresses foam cell formation by activating AMP activated protein kinase (AMPK) pathway and inhibiting NF $\kappa$ B/NLRP3 signaling axis, thus inhibiting macrophage polarization into inflammatory cells and limiting the uptake of oxLDL [23]. Previous studies on the *S. alba* fruit extract have demonstrated its capacity to suppress LDL-C and triglyceride (TG) as well as to reduce the atherogenic index plasma *in vivo* [24].

Additionally, the *S. alba* fruit extract inhibits foam cells formation, prevents microvascular structural alterations toward atherosclerosis and diminishes tumor growth factor- $\beta$  (TGF- $\beta$ ) expression in high fat diet animal model [24,25]. *In silico* analysis indicates that *S. alba* fruit possesses active metabolites that can interact with three ROS proteins through hydrogen bonding [27].

The existing literatures, however, remain limited to the phytochemical compounds and antioxidant capacity of *S. alba* fruit. In our knowledge, there is no research has thoroughly investigated the potential of *S. alba* fruit extract in preventing atherogenesis, particularly its effects on LOX-1 expression, as well as the cellular antioxidant response mediated by Nrf2 and its downstream antioxidant MnSOD. This research examined the effect of *S. alba* fruit extract on atherosclerosis progression in high-fat diet-induced rat model, focusing on the alterations in the aortic arch. Spastically we aimed to determine whether the extract could modulate the expression of the pro-atherogenic receptor LOX-1 and its downstream consequence, foam cell formation. Furthermore, we sought to elucidate the expression of the critical antioxidant transcription factor Nrf2 and one of its key target enzymes, MnSOD. We hypothesized that *S. alba* fruit extract could inhibit early atherogenesis effect by concurrently reducing LOX-1 expression and elevating the Nrf2 mediated antioxidant defense system.

## Materials and methods

### Study design and ethical approval

This study performed a post-test control only group design using experimental animals to evaluate the effect of *S. alba* fruit extract. The research strictly adhered to the ethical guidelines for the animal experimentation. Ethical approval for the study was obtained from Research Ethical Committee of the Faculty of Medicine, University of Andalas, Padang, Indonesia under approval number 231/UN.16.2/KEP-FK/2024.

### Preparation of *Sonneratia alba* fruit extract

Unripe fruit of *Sonneratia alba* measuring 2 - 3 cm in diameters were collected from the coastal area of Sungai Apit Regency, Siak Sri Indrapura District, Riau Province, Indonesia. The species was authenticated by a botanist from Faculty of Mathematic and Natural

Sciences, University of Riau, Pekanbaru, Indonesia with voucher specimen number 583/UN19.5.1.1.3-4.1/TU.00.01/2025. After removing its calyces, the fruits were thoroughly washed, oven-dried at a degree of 40 - 45 °C, and ground into a coarse powder. From 4 kg fruit, approximately 670 g crude powder were obtained, corresponding to an estimated yield of 16.75%. The powdered material was subjected to maceration by 96% ethanol (1 gram per 10 mL solvent) for 72 h, with intermittent stirring. The solvent was then filtered and stored in a dark room at room temperature. The maceration was done twice. The filtrate was concentrated *in vacuo* using a rotary evaporator at 40 °C to yield the crude ethanol extract. The extract was stored in refrigerator until used. Qualitative phytochemical analysis of the *S. alba* fruit extract confirmed the presence of flavonoids, phenolics, saponins and steroids. The extract was further quantified for its secondary metabolites. The total phenolic content (TPC), determined using the Folin-Ciocalteu assay [16], was measured at  $171.99 \pm 1.53$  mg GAE/g extract. The total flavonoid content (TFC), assessed by the aluminum chloride colorimetric method [16], was found to be  $14.49 \pm 0.31$  mg QE/g extract. The antioxidant capacity of the extract was evaluated using the DPPH (2,2-diphenyl-1-picrylhydrazyl) radical inhibition assay, yielding an IC<sub>50</sub> value of 61.42 ppm. This result indicate that the extract possesses strong antioxidant activity.

### Experimental animal and atherogenesis induction

A total of 24 male Wistar rats (*Rattus norvegicus*), aged 8 - 10 weeks and weighing 170 - 190 g, were procured from the Faculty of Pharmacy, University of Andalas, Padang. The sample size was determined using the Resource Equation method. The animals were housed under standard laboratory condition with a 12-hours light/dark cycle,  $22 \pm 2$  °C and *ad libitum* access to water.

Atherosclerosis was induced by feeding rats a high fat diet (HFD) of 20 g per day. This HFD protocol was adapted from Ismawati *et al.* [28] with two primary modifications; the high-dose administration of vitamin D3 (700.000 IU/kg) was omitted to avoid the masking effect of high-dose vitamin D3 effect on protein targets expression, and the diet duration was extended to 10 weeks, following the established publication to induce

hyperlipidemia and early atherogenesis with high fat diet in rats [29,30]. This modified protocol was established through a preliminary study to optimize and confirm the diet duration and effectiveness in reliably inducing early atherogenesis in rats, characterized by significant LDL-C elevation and observable foam cell accumulation in aortic arch of rat model (data not shown). The HFD formulation consisted of standard chow Vivo512 (Charoen Pokphand Indonesia Tbk, Medan, Indonesia) supplemented by 2% cholesterol-derived from egg yolk, 5% goat fat, and 0.2% cholic acid (Sigma Aldrich, St. Louis, USA). The feeding and treatment protocols were conducted for a total duration of 10 weeks.

Rats were randomly divided into six group (n = 4 per group); normal control (NC) fed standard chow Vivo512; high-fat diet control (HFD-C) fed HFD; high-fat diet + atorvastatin (HFD-A) fed HFD and administrated atorvastatin (KalbeMED, Jakarta, Indonesia) at a converted dose of 10 mg/day by oral gavage; high-fat diet + extract 200 (HFD-E200) fed HFD and administrated *S. alba* fruit extract at a dose of 200 mg/kg body weight/day by oral gavage; high-fat diet + extract 400 (HFD-E400) fed HFD and administrated *S. alba* fruit extract at a dose of 400 mg/kg body weight/day by oral gavage; and high-fat diet + extract 800 (HFD-E800) fed HFD and administrated *S. alba* fruit extract at a dose of 800 mg/kg body weight/day by oral gavage.

At the end of the 10-week treatment, the animals were fasted overnight and then deeply anesthetized using an intra peritoneal injection of ketamine (70 mg/kg) and xylazine (0.7 mg/kg). Blood samples were collected via cardiac puncture for LDL-C and HDL-C plasma levels analysis. Following perfusion with normal saline to remove residual blood, the aortic arch was carefully excised from each rat and immediately fixed in 10% neutral buffered formalin for subsequent histological and immunohistochemical analysis.

#### **Histological and immunohistochemical analysis**

Fixed aortic arch tissues were routinely processed and embedded in paraffin blocks. Section of 3  $\mu$ m thickness were cut and stained with hematoxylin and eosin (H&E) to appraise general morphology and detect foam cells. The foam cells were identified in the intimal and medial layer of aortic arch by using a light

microscope (Leica IC550HD, Leica Mycosystem, Danaher Life Sciences, Germany). Photomicrographs were captured in all microscopic fields of 400x magnification by using Leica Application Suite Las Ez v3.4.0 (build:272) (Leica Mycosystem, Switzerland) and the foam cells were quantified by using ImageJ v1.54g software (Wayne Rasband, National Institutes of Health, USA).

Immunohistochemistry was performed on 3  $\mu$ m paraffin-embedded aortic arch section to assess the expression indexes of LOX-1, Nrf2 and MnSOD. Briefly, the sections were deparaffinized, rehydrated and subjected to antigen retrieval using a citrate buffer. Endogenous peroxidase activity was quenched with 3% H<sub>2</sub>O<sub>2</sub>. The sections were then incubated for one hour at room temperature with primary antibodies anti-LOX-1 1:50 (Cat PA5-102452, Thermo Fisher Scientific, USA), anti-Nrf2 1:50 (SAB4501984, Sigma-Aldrich, Darmstadt, Germany) and anti-MnSOD 1:300 (BT Lab, Zhijiang, China). Following washing, an appropriate biotinylated secondary antibody and a streptavidin-HRP complex were applied. Immunoreactivity was visualized using 3,3'-diaminobenzidine (DAB) substrate, resulting in brown precipitates at the site of antigen binding. The sections were then counterstained with Mayer's hematoxylin.

The expression indexes of LOX-1, Nrf2, and MnSOD were quantified using Colour Deconvolution v2 plugin integrated into ImageJ v1.54g software, across eight random high-power field photomicrographs (400x magnification). The expression index was calculated as  $\Sigma$  (intensity x percentage of positive cells). The intensity was obtained from mean grey value and the percentage of positive cells were obtained from percentage area measured in ImageJ.

#### **Statistical analysis**

All quantitative data were expressed as the mean and Standard Error of Mean (SEM). The normality of the data distribution was assessed using the Saphiro-Wilk test. Differences among the six groups were analyzed using One-Way Analysis of Variance (ANOVA) or Kruskal Wallis-test, followed by Tukey's Post Hoc test or Dunn's test for multiple pairwise comparisons. A *p* - value of less than 0.05 was considered statistically significant. All statistical

analysis was performed using Graphpad Prism v9.0.0 software (Graphpad Software, Boston, MA, USA).

## Results and discussion

### Effect of *Sonneratia alba* fruit extract on plasma lipid profile

The impact of the high-fat diet and the subsequent treatment with *S. alba* fruit extract on plasma low-density lipoprotein cholesterol (LDL-C) and high-density lipoprotein cholesterol (HDL-C) levels is presented in **Table 1**. The HFD regimen effectively induced hypercholesterolemia as demonstrated by a highly significant increase in plasma LDL-C level relative to the normal control (NC) group (~3.3 fold increase,  $p < 0.05$ ), confirming the successful induction of hyperlipidemia and the pro-atherogenic state. Treatment with a high dose of *S. alba* fruit extract (HFD-E800) significantly showed a potent hypolipidemic effect, achieving a statistically significant reduction in LDL-C that nearly normalizing the levels to those of the NC group. Moreover, the efficacy of the 800 mg/kg/day

extract dose was comparable to the positive control, Atorvastatin (HFD-A). This indicates that the extract exerts a significant potential to modulate this key atherogenic risk factor. Conversely, the lower dosages of *S. alba* (HFD-E200 and HFD-E400) exhibited a modest, dose-dependent tendency towards reduction, while the difference was not statistically significant compared to HFD-C.

In contrast to LDL-C, the concentrations of HDL-C plasma levels showed no statistically significant difference across all six experimental groups. The HFD-C group had a minimal, non-significant reduction compared to the NC group. Treatment with *S. alba* extract, even at the highest dosage, did not significantly restore or modulate this cardio-protective lipid fraction. This finding indicates that the primary hypolipidemic effect of the *S. alba* fruit extract is mediated through the diminishing of atherogenic lipoproteins (LDL-C) rather than by the augmenting of cardio-protective lipoproteins (HDL-C).

**Table 1** Plasma LDL-C and HDL-C levels across experimental groups (means  $\pm$  SEM).

Group	LDL-C (mg/dL) $\pm$ SEM	HDL-C (mg/dL) $\pm$ SEM
Normal control (NC)	46.63 $\pm$ 5.41 <sup>###</sup>	51.30 $\pm$ 3.02
HFD control (HFD-C)	155.40 $\pm$ 14.61	46.90 $\pm$ 4.95
HFD + atorvastatin (HFD-A)	81.00 $\pm$ 16.35 <sup>#</sup>	45.35 $\pm$ 1.75
HFD + extract 200 mg (HFD-E200)	101.20 $\pm$ 7.39	47.00 $\pm$ 3.62
HFD + extract 400 mg (HFD-E400)	94.68 $\pm$ 23.31	46.50 $\pm$ 4.18
HFD + extract 800 mg (HFD-E800)	59.38 $\pm$ 4.67 <sup>##</sup>	49.15 $\pm$ 1.30
<i>p</i> - value	0.0005*	0.7509

\*indicates a significant different across the experimental groups ( $p < 0.05$ ). #, ## and ### indicate a significant different compared to HFD-C group with  $p < 0.05$ ,  $p < 0.01$  and  $p < 0.001$ , respectively.

The significant decrease in LDL-C plasma levels, particularly by the 800 mg/kg/day dose of *S. alba* fruit extract is a crucial finding as hypercholesterolemia is the fundamental risk factor for atherogenesis initiation. This preliminary observation highlights the extract's potential as a natural agent for controlling hyperlipidemia. The mechanism responsible for this hypolipidemic effect remains hypothetical, as specific biochemical assays such as enzyme activity or receptor quantification were beyond the scope of this study. The hypolipidemic effect may be associated with the

phytoconstituents such as polyphenol, flavonoid, sterols and triterpenoids previously identified in *Sonneratia* species [16,17]. From mechanistic point of view, phytocompounds may interfere with the cholesterol absorption in the intestine, possible by binding to bile acids and thus increase fecal excretion, modulate key enzymes in hepatic lipogenesis, such as 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase that is functioning similarly to statins like Atorvastatin. Subsequently, the extract may facilitate to enhance circulating lipids catabolism by modulating hepatic

LDL receptor (LDL-R) expression, therefore augmenting the clearance of LDL-C from the plasma or boosting cholesterol-7 $\alpha$ -hydroxylase (CYP7A1) enzyme involved in converting cholesterol into bile acids [31-33]. Polyphenols reduce LDL-C level by enhancing the activity of liver X receptor  $\alpha$  (LXR $\alpha$ ), suppressing hepatocyte nuclear factor 1  $\alpha$  (HNF-1 $\alpha$ ), consequently diminishing the mRNA expression of proprotein convertase subtilisin/kexin type 9 (PCSK9). This mechanism ultimately results in increased LDLR expression and enhanced LDL-C uptake [34]. These pathways suggest a direct effect on the clearance of LDL-C without necessarily affecting the synthesis or reverse cholesterol transport function of HDL-C as observed in a hypercholesterolemic male rabbits fed 0.2% curcumin [35].

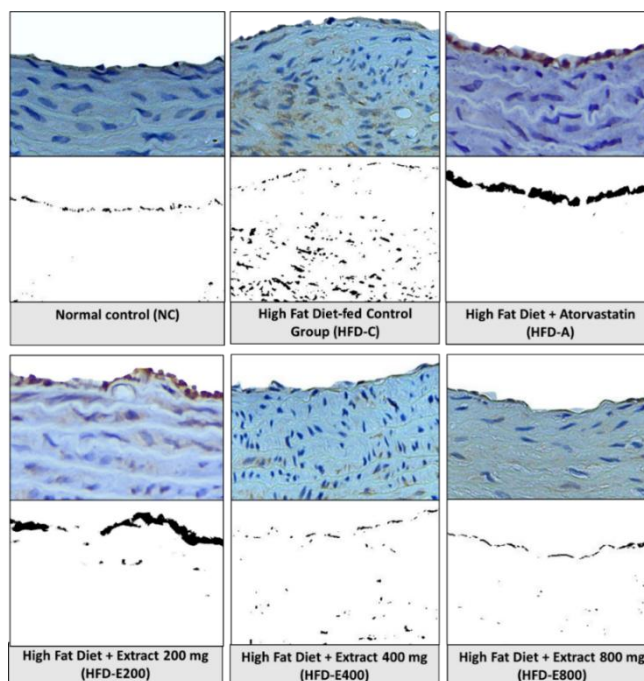
The lack of significant effect of HDL might be explained by the high complexity of HDL metabolism. However, this result is consistent with the existing hyperlipidemia therapies, in which high-efficacy therapeutic classes such as statin and cholesterol absorption inhibitors provide major clinical benefits despite minimal HDL-C augmentation [36]. The metabolism of HDL-C involves various regulatory steps, including the function of cholesteryl ester transfer protein (CETP), lecithin-cholesterol acyltransferase (LCAT) and the expression of ATP-binding cassette transporter A1 (ABCA1) for cholesterol binding. The phytochemicals do not seem to strongly interfere with this specific enzymes or transporters that control HDL [37]. Elevated plasma HDL concentration is not a reliable surrogate for low hyperlipidemia risk, instead, it may reflect increased HDL turnover or functional demand in response to high circulating LDL-C burden. Consequently, determining atherosclerosis risk merely by quantifying HDL-C concentration is insufficient

[38]. Further investigation is warranted to assess HDL functional capacity, such as its cholesterol efflux capability. It will determine whether the extract preserves the qualitative protective properties of HDL even without altering its plasma concentration.

The significant decrease in LDL-C levels necessitates an examination of the downstream events occurring within the arterial wall. The essential process in atherogenesis is the transformation of LDL into oxidized LDL which serve as the primary ligand for LOX-1 receptor and the key trigger for the inflammatory cascade [39]. Therefore, we hypothesized that the extract efficacy in reducing LDL-C would result in a tangible reduction in vascular pathology. The expression of pro-atherogenic receptor LOX-1 and its downstream consequent foam cell formation are discussed in the following section.

#### **LOX-1 expression and foam cell formation in aortic arch**

Lectin-like oxidized low-density lipoprotein receptor 1 (LOX-1) is a key molecular driver of early atherogenesis, and its expression in the aortic arch of rats was analyzed using immunohistochemical staining (**Figure 1**). In the normal control (NC) group, minimal LOX-1-expression was detected. In contrast, the high-fat diet control (HFD-C) group exhibited a pronounced and diffuse upregulation of LOX-1 expression, predominantly localized in the endothelial cells lining the intima and subendothelial cells likely infiltrating macrophages and/or transdifferentiated vascular smooth muscles (VSMCs). This confirmed the pathological activation of the pro-atherogenic state induced by HFD. Quantitative analysis (**Table 2**) confirmed that the HFD-C group displayed a more than threefold increase in LOX-1 expression compared to NC group.



**Figure 1** Immunohistochemical analysis of LOX-1 expression in the aortic arches of experimental groups. Representative images of immunohistochemical staining for LOX-1 (brown chromogen) in the intimal and medial layers of the rat aortic arch (top row of each panel) and the corresponding processed images highlighting positive staining using Colour Deconvolution 2 plugin integrated in ImageJ software (bottom row of each panel) are revealed at 400x magnification.

Treatment with *Sonneratia alba* fruit extract demonstrated a clear, dose-dependent decreasing in LOX-1 expression, with significant difference observed across the experimental group (**Table 2**). The HFD-E400 group showed a significant reduction of LOX-1 expression compared to the HFD-C group. The HFD-E800 group achieved the most pronounced attenuation, approximately 63% reduction compared to HFD-C group, resulting in a statistically significant reduction

that returned LOX-1 expression to levels closely resembling those of the NC group. This indicates a powerful suppression effect of *S. alba* extract on the expression of LOX-1 receptor that is responsible for oxLDL uptake. Although the positive control (HFD-A) also showed a reduction in LOX-1 expression, however, its effect was not statistically significant compared to HFD-C group (**Table 2**).

**Table 2** Expression index of LOX-1 and foam cell number in aortic arch of experimental groups.

Group	LOX-1 expression index $\pm$ SEM	Foam cell number $\pm$ SEM
Normal control (NC)	51.08 $\pm$ 2.32	275.5 $\pm$ 123.9
HFD control (HFD-C)	160.30 $\pm$ 19.03	932.5 $\pm$ 238.5
HFD + atorvastatin (HFD-A)	105.20 $\pm$ 21.08	481 $\pm$ 188.9
HFD + extract 200 mg (HFD-E200)	94.15 $\pm$ 11.14	433 $\pm$ 78.52
HFD + extract 400 mg (HFD-E400)	83.43 $\pm$ 20.95 <sup>#</sup>	358 $\pm$ 162.4
HFD + extract 800 mg (HFD-E800)	58.96 $\pm$ 15.80 <sup>##</sup>	168 $\pm$ 42.44 <sup>#</sup>
<i>p</i> - value	0.0051*	0.0398*

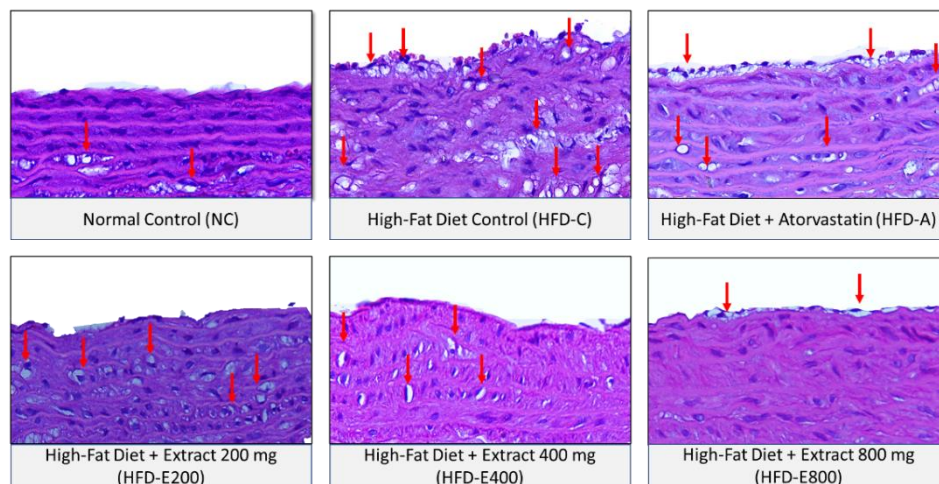
\*indicates a significant different across the experimental groups ( $p < 0.05$ ). # and ## indicate a significant different compared to HFD-C group with  $p < 0.05$  and  $p < 0.01$ , respectively.

The histological quantification of lipid-laden cells or foam cells in the aortic arch supported the molecular findings regarding reduction of LOX-1 expression (Table 2). The HFD-C group demonstrated the highest accumulation of foam cells, serving as a definitive morphological indicator of early atherogenic lesions in the model. Histological examination revealed these cells distributed across both the intimal and medial layers of the aorta in HFD groups (Figure 2), suggesting participation from both infiltrating macrophages and trans-differentiated vascular smooth muscle cells (VSMCs) in the complex lesion morphology.

The administration of *S. alba* fruit extract resulted in a significant and dose-dependent reduction of foam cells formation ( $p = 0.0398$ ). The highest dosage group, HFD-E800, exhibited the most significant effect on reducing foam cells accumulation. This reduction was statistically significant compared to the HFD-C group ( $p = 0.0256$ ), demonstrating a remarkable anti-atherogenic influence. The HFD-E400 group also exhibited a prominent reduction, outperforming the atorvastatin group, however, this reduction was not statistically

significant compared to HFD-C. Moreover, the expression of LOX-1 significantly correlated with accumulation of foam cells ( $p = 0.0005$ ,  $R^2 = 0.4462$ ), suggesting this direct morphological evidence confirms that the extract interferes the pathological uptake and retention of lipids by vascular cells.

The quantity of foam cells observed in this study has a high variability in almost all groups. This observed variability may be a reflection of biological heterogeneity to this specific *in vivo* model of early atherogenesis, which is commonly reported in diet-induced rodent models. This variation reflects the heterogeneous metabolic and inflammatory responses of individual rats to the chronic HFD-induced stress and hyperlipidemia [38]. Moreover, foam cells are the manifestation of highly localized and scattered early lesions or fatty streaks [40]. By analyzing a limited number of tissue sections, in this case aortic arch, inherent non-uniformity in lesion distribution throughout the entire aortic tree in evitable contributes to a larger standard deviation.



**Figure 2** Histopathological analysis of foam cells formation in the aortic arch of experimental groups (Hematoxylin and Eosin staining). Representative image of aortic arch section stained with H&E at 400x magnification. Red arrows denote lipid-laden foam cells within the intimal and medial layers of the vascular wall.

Lectin like oxidized low-density lipoprotein receptor 1 (LOX-1) is a key scavenger receptor for oxLDL, distinct from CD36 in its recognition of Apolipoprotein-B modification on oxLDL rather than oxidized phospholipid [6,41]. While normally expressed at low levels in monocytes, LOX-1 expression is markedly upregulated by pro-atherogenic stimuli such

as oxLDL, high glucose and pro-inflammatory cytokines. In addition to monocytes and macrophages, LOX-1 is expressed by endothelial cells and VSMCs [39], aligning with its pervasive role in arterial wall pathology. The upregulation of LOX-1 activates the NF- $\kappa$ B pathway, a crucial signaling cascade involved in multiple stages of atherogenesis, from plaque formation,

instability and rupture. NF- $\kappa$ B not only regulates the expression of several pro-inflammatory mediators that exacerbate atherosclerosis but also establishes a positive feedback loop by directly enhancing LOX-1 expression through binding to its promoter [42,43]. Consequently, the inhibition of LOX-1 expression is a pivotal intervention against this vicious cycle of inflammation and lipid accumulation [7].

The HFD model effectively replicates early atherogenesis, annotated by hypercholesterolemia and the subsequent overexpression of LOX-1. The significant overexpression of LOX-1 in the HFD-C group provides a molecular link that may explain the massive accumulation of foam cells. LOX-1 activation accelerates oxLDL uptake, leading to the uncontrolled formation of lipid-laden macrophages and VSMCs, that constitute the fundamental structural components of fatty streak [39].

The significant reduction of LOX-1 expression by the 800 mg/kg/day dose of *S. alba* fruit extract suggests a directly correlation with the remarkable 82% reduction in the foam cell numbers compared to the HFD-C group. This strong correlation suggests that the *S. alba* effect in preventing early atherogenesis is, at least in part, a direct consequence of down-regulating this key scavenger receptor. This molecular influence is probably mediated by the extract's ability to lower plasma LDL-C, thereby reducing oxLDL substrate, and by its inherent antioxidant properties that limit oxLDL formation, collectively diminishing the stimuli required for LOX-1 gene activation.

The enhanced efficacy of HFD-E800 compared to the atorvastatin group in decreasing LOX-1 expression and foam cell generation is noteworthy. Statins, such as atorvastatin, are primarily recognized for their capacity to lower circulating LDL-C; nevertheless, they also exert pleiotropic effects, including certain anti-inflammatory and antioxidant activities [44]. The *S. alba* extract, characterized by its intricate variety of phytocompounds, seems to present a more effective multi-target intervention. The extract likely achieves enhanced suppression of the LOX-1 expression, either independently or in addition to its influence on circulating lipids.

The significant capacity of *S. alba* fruit extract to reduce LOX-1 expression is probably due to its rich content of secondary metabolites. These compounds are

hypothesized to act through dual mechanisms; reducing circulating oxLDL levels, thereby dampening the primary stimulus for LOX-1 activation, and directly inhibiting NF- $\kappa$ B activation, that prevents its binding to the LOX-1 promoter and consequently suppressing LOX-1 transcription [45]. These findings are supported by previous researches on similar natural compound. Flavonoids found in hawthorn leaves (e.g., luteolin and vitexin, also found in *S. alba*) suppress LOX-1 expression in macrophages, partly through inhibition of the SCAP-SREBP2-LDLR pathway [46]. Oleanolic acid, a secondary metabolite found in *S. alba* fruits [47,48], significantly inhibited oxLDL-stimulated LOX-1 expression in HUVEC cells [19]. Curcumin, bergamot peel, ellagic acid and quercetin diminish oxLDL-induced ROS production and suppressing LOX-1 expression and lipid accumulation in macrophages, frequently by modulating signaling pathway such as signal transducer and activator transcription 3 (STAT3) or reducing endothelial reticulum (ER) stress [49]. In addition to direct receptor modulation, natural agents hydroxytyrosol and chitosan oligosaccharide inhibit foam cells formation by stimulating cholesterol efflux and suppressing influx, particularly via PPAR $\gamma$ /LXR $\alpha$  signaling pathway and subsequent upregulation of ABCA1 [50,51]. These findings jointly suggest the potential of *S. alba* phytochemicals to directly modulate LOX-1 expression.

The histological observation that foam cells originated from both macrophages and VSMCs underscores the complexity of the lesions. The ability of the *S. alba* extract to effectively reduce the formation of these cells suggests a protective action extending to both cell types, suggesting a profound ability to stabilize the vessel wall environment. Recent evidence highlights the significant contribution of VSMC to the foam cell population, with studies suggesting VSMC-derived foam cell can account for a substantial portion of the lipid-laden cells in plaques [52,53]. These VSMC-derived foam cells are often less efficient at eliminating lipids, contributing significantly to disease progression and plaque instability [52,54].

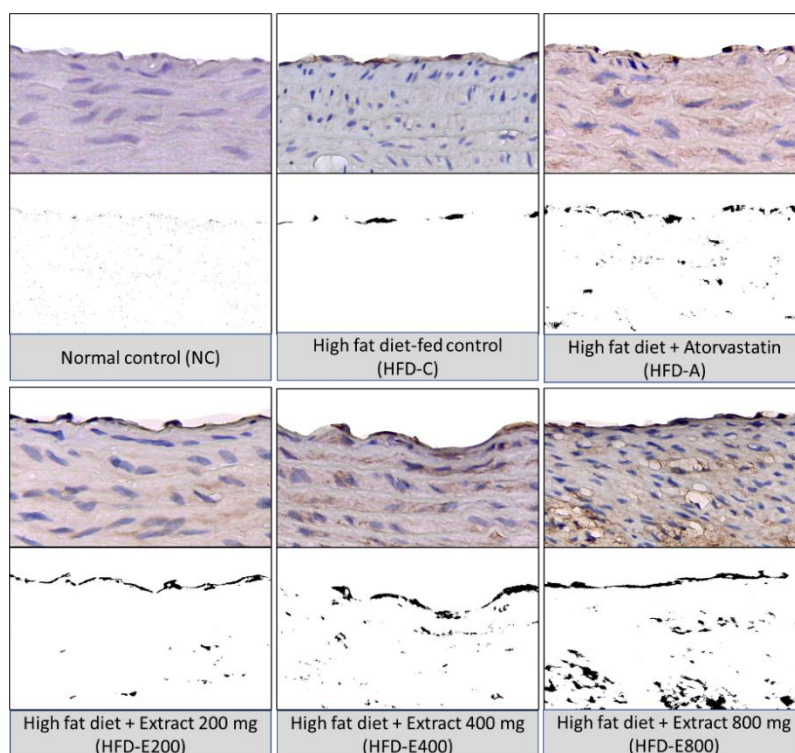
In the present study, the presence of foam cells in both the intimal and medial layers of H&E-stained aortic arch sections strongly suggests that VSMC-derived foam cells likely constitute a substantial, if not dominant, portion of the lipid-laden cells observed in

our HFD model. Future studies are warranted to precisely uncover the origin of foam cells in this model through specific markers (e.g.,  $\alpha$ -smooth muscle actin for VSMCs- and CD68 for macrophages-derived foam cells).

### Expression of Nrf2 and MnSOD in aortic arch of rats

Nuclear factor erythroid-2 related factor 2 (Nrf2) immunoreactivity in the normal control (NC) group was observed at low basal levels, predominantly within the cytoplasm of endothelial cells and scattered subendothelial cells. The high-fat diet control (HFD-C)

group exhibited a marked increase in overall Nrf2 expression compared to NC group, suggesting an attempted compensatory response to elevated oxidative stress. Treatment with *S. alba* fruit extract resulted in a profound and dose-dependent upregulation of Nrf2 expression. The HFD-E800 group displayed the most intense Nrf2 immunoreactivity, characterized by widespread Nrf2 expression within the endothelial cells lining the lumen and in the subendothelial cellular populations, likely macrophages and VSMCs. The HFD + Atorvastatin (HFD-A) group also exhibited an increase in Nrf2 expression, particularly in nuclear staining. (Figure 3).



**Figure 3** Immunohistochemical analysis of Nrf2 expression in the aortic arch of experimental groups. Representative images of immunohistochemical staining for Nrf2 in the intimal and medial layers of the rat aortic arch (top row of each panel) and corresponding processed images highlighting positive staining (bottom row of each panel) are shown at 400x magnification.

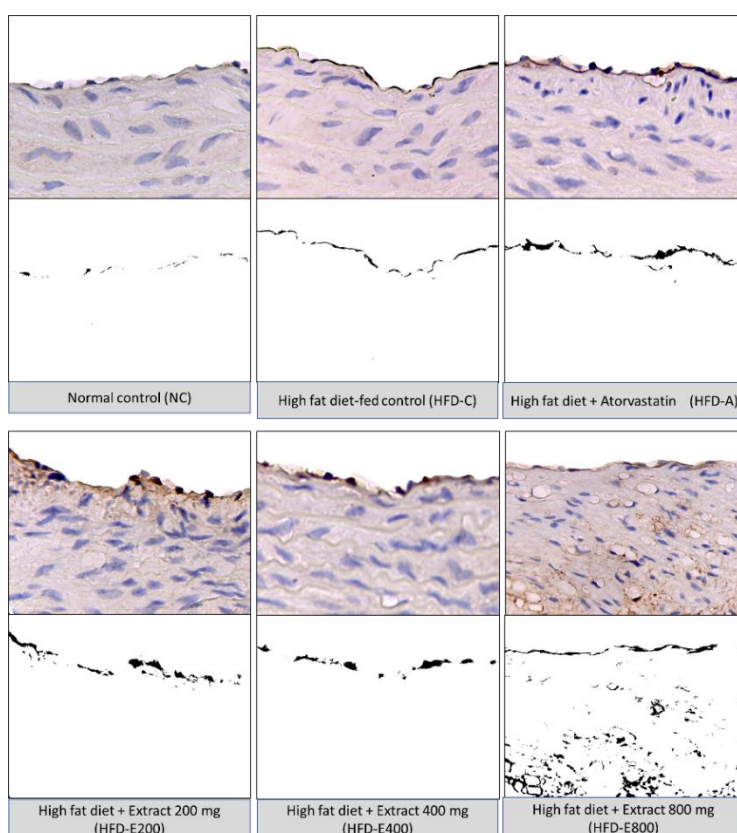
The expression index analysis of the Nrf2 in the aortic arch was significantly modulated by the treatments ( $p = 0.0059$ ). The *S. alba* extract elevated Nrf2 expression in which the HFD-E800 group

exhibited the highest Nrf2 expression index, that was significantly higher than that of HFD-C ( $p = 0.0486$ ) (Table 3).

**Table 3** Expression index of Nrf2 and MnSOD in aortic arch of experimental groups.

Group	Nrf2 Expression Index $\pm$ SEM	MnSOD Expression Index $\pm$ SEM
Normal control (NC)	35.39 $\pm$ 6.22	48.29 $\pm$ 4.58
HFD control (HFD-C)	91.57 $\pm$ 10.76	64.62 $\pm$ 14.64
HFD + atorvastatin (HFD-A)	149.70 $\pm$ 10.71	103.20 $\pm$ 12.64
HFD + extract 200 mg (HFD-E200)	144.40 $\pm$ 32.75	116.50 $\pm$ 30.44
HFD + extract 400 mg (HFD-E400)	197.10 $\pm$ 49.46	106.30 $\pm$ 18.45
HFD + extract 800 mg (HFD-E800)	224.40 $\pm$ 32.73 <sup>#</sup>	144.90 $\pm$ 22.54
<i>p</i> -value	0.0059*	0.0433*

\*indicates a significant different across the experimental groups ( $p < 0.05$ ). <sup>#</sup> indicates a significant different compared to HFD-C group with  $p < 0.05$ .



**Figure 4** Immunohistochemical analysis of MnSOD expression in the aortic arch of experimental groups. The top row of each panel represents images of immunohistochemical staining for MnSOD in the intimal and medial layers of aortic arch of rats, while the bottom row of each panel represents corresponding processed images highlighting positive staining. The figures are shown at 400x magnification.

The expression of MnSOD, a down-stream target of the Nrf2 pathway, was analyzed by using immunohistochemistry assay (**Figure 4**). The normal control (NC) group displayed MnSOD immunoreactivity at low basal levels within cytoplasm of endothelial and scattered subendothelial cells. In contrast, the high-fat diet control (HFD-C) group

exhibited a marked increase in MnSOD expression compared to NC, suggesting a compensatory upregulation in response to increase oxidative stress associated with the HFD. Treatment with *S. alba* fruit extract demonstrated a dose-dependent enhancement of MnSOD expression compared to HFD-C. The HFD-E800 group exhibited the highest MnSOD

immunoreactivity with strong cytoplasmic granular staining prominently in the endothelial cells and extending into subendothelial cellular populations. However, despite this apparent dose-dependent increase, the enhancement in MnSOD expression across all treatment groups did not reach statistical significance compared to the HFD-C group (**Table 3**). This finding indicates that while the extract may influence the expression of antioxidant enzymes, the effect on MnSOD abundance was not definitively confirmed by this analysis.

The upregulation of Nrf2 protein levels represents a key molecular correlate to the observed anti-atherosclerotic effects of the *S. alba* extract. Cellular oxidative or electrophilic stress fundamentally disrupts intracellular redox balance, necessitating an adaptive response [55]. The Keap1-Nrf2 system serve as the principal cytoprotective mechanism, regulating over 1,000 genes involved in inflammation and lipid metabolism [56].

The elevated Nrf2 protein expression achieved by the *S. alba* extract is highly relevant, given that Nrf2 expression is often suppressed in chronic conditions like diabetes, hypertension and inflammation [57]. The enhanced antioxidant defense is hypothesized to reduce oxLDL formation by neutralizing reactive oxygen species (ROS) [58], thereby preventing the oxidation of circulating LDL-C and suppressing LOX-1 expression. Decreased cellular oxidative stress is known to downregulate LOX-1 gene expression [39]. The anti-atherogenic role of Nrf2 is supported by genetic evidence where deletion of Nrf2 in LDLR<sup>-/-</sup> mice exacerbates atherosclerotic lesions and upregulates pro-inflammatory genes expression [59]. Following this proposed mechanism, the extract likely performs a coordinated protective action, blocking the LDL substrate via its hypolipidemic effect and suppressing the oxLDL receptor by augmenting the Nrf2-driven antioxidant defense.

The observed Nrf2 expression elevation is attributed to the rich flavonoid and polyphenol content of *S. alba* fruit. These phytochemical compounds are known to activate Nrf2 by altering cysteine residues (e.g., Cys151) on Keap1, thereby disrupting the Keap1-Nrf2 complex and facilitating the nuclear translocation and gene transcription of Nrf2 [60]. This finding are supported by previous studies demonstrating that the

plant-derived compounds induce Nrf2 activity, subsequently triggering multiple pathways that preserve mitochondrial integrity and enhanced metabolic activity [12]. The high clinical and genetic relevance of Nrf2 reinforces its status as a critical target in mitigating vascular disease [61,62].

The analysis of MnSOD, a key mitochondrial antioxidant enzyme, revealed an increase in expression reaching its peak in the HFD-E800 group. However, this increase was not statistically significant compared to the HFD-C group (**Table 3**). This non-significant result may reflect a decoupling phenomenon, where the upregulation of Nrf2 protein preferentially drives the expression of other antioxidant enzymes (e.g. HO-1, NQO1) or is subjected to post transcriptional control that limits the measurable increase in MnSOD [63].

MnSOD is essential for reducing mitochondrial damage and its expression is directly associated with anti-apoptotic defense within atherosclerotic lesions [64,65]. While the non-significant change limits definitive conclusion, the observed trend aligns with the mechanism of action reported for similar phytochemical such as luteolin and baicalein which augmented SOD expression via Nrf2 or related pathway [14]. Nevertheless, the upregulation of the master regulator Nrf2 protein expression remains the most significant finding in elucidating the extract potential for vascular protection. Definitive confirmation of Nrf2 activation and its impact on downstream enzymatic activity requires functional assays, such as nuclear-cytoplasmic translocation and MnSOD activity assay, which were beyond the scope of this study.

## Conclusions

This study demonstrates that the ethanolic fruit extract of *S. alba* effectively ameliorates key early atherogenic features in rats fed a high fat diet-induced rat model, showing a strong association with several changes in both circulating lipids and vascular markers. The optimal dose of 800 mg/kg/day was associated with a significant reduction of plasma LDL-C levels, a primary precursor to plaque formation. Within the aortic arch, the extract demonstrated a coordinated protection action, leading to a significant reduction of the pro-atherogenic receptor LOX-1 and which correlated with a marked reduction in foam cell accumulation. This protective effect is further molecularly associated with

an upregulation of the cytoprotective protein Nrf2 expression, suggesting an influence on the endogenous antioxidant defense system. While the enhancement of the downstream enzyme MnSOD did not reach statistical significance, the combine effect of Nrf2 elevation and the concurrent reduction of LOX-1 expression and foam cell formation suggests that *S. alba* may serve as a potential intervention strategy by targeting hyperlipidemia and modulating oxidative stress markers for the initiation of atherogenesis. However, due to the limited scope of the current findings, which focused primarily on protein expression level and surrogate markers, further functional studies, such as nuclear translocation assays, enzymatic activity and definitive receptor binding, are warranted to confirm its proposed multi-target mechanism and overall long-term anti-atherosclerotic efficacy.

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#### Declaration of Generative AI in Scientific Writing

The authors declare that certain generative AI tools such as QuillBolt and Grammarly were utilized solely during the preparation of this manuscript for language refinement, grammar correction and style editing. No part of the scientific content, research findings, data interpretation or conclusion was generated by any AI tool. The authors affirm complete responsibility for the entirety of the work presented herein and the conclusion drawn.

#### CRedit Author Statement

**Huriatul Masdar:** Conceptualization, Methodology, Software, Formal analysis, Investigation, Validation, Visualization, Funding acquisition, and Writing original draft. **Nur Indrawaty Lipoeto:** Validation, Data curation, Review and Editing, Project administration and Supervision. **Gusti Revilla:** Methodology, Validation, Investigation, Data Curation, Review and Editing, and Supervision. **Eka Fithra Elfi:** Validation, Software, Formal analysis, Data Curation,

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